RESEARCH PAPER



THE EFFECT OF L-ORNITHINE ON THE INNER EAR OF RATS IN THE LIPOPOLYSACCHARIDE INDUCED ENDOTOXEMIA MODEL

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SUMMARY

Objectives; To investigate the relationship between ornithine treatment, p53 expression and the histopathologic differentials of rats' inner ear in endotoxemia induced by lipopolysaccharide. Material and methods; Twentyfour healthy rats were randomly divided into 4 groups. The first group, served as control, received intraperitoneal saline; the second group, intraperitoneal endotoxin; the third group, intraperitoneal ornithine and saline, and the fourth group received intraperitoneal ornithine ten minutes before endotoxin administration. Results; Congestion and subepitelial edema were observed in stria vascularis, and leukocyte, plasma cell infiltration in the parts of membraneuos labyrinthitis of modiolar region in the cochlea of the second and the fourth group. Inflammation was not detected in the parts of vestibulocochlear and facial nerves in meatus acousticus internus. When we compared the plasma levels of nitrite/nitrate and MDA among groups, we found statistically significant differences: between groups 1 and 2, 1 and 3, 1 and 4, 2 and 3, 3 and 4. However, no significant difference was found in groups 2 and 4. Ornithine did not reduce the harmful effect of sepsis in rats. Immunohistochemically, p53 immunoreactive cells were not detected in any groups. Conclusion; L-ornithine can not improve the manifestations of septic picture in the rat cochlea.

Keywords: Inner ear immunity, labyrinthitis, sepsis, endotoxin, hearing loss, endotoxemia

LİPOPOLİSAKKARİT İLE OLUŞTURULAN ENDOTOKSEMİ MODELİNDE L-ORNİTİNİN RAT İÇ KULAĞINA ETKİSİ

ÖZET

Amaç: Lipopolisakkarit ile oluşturulan endotoksemide rat iç kulak histopatolojisi ile ornitin tedavisi ve P 53 ekspresyonu arasındaki ilişkinin araştırılması. Gereç ve Yöntemler: Yirmi dört sağlıklı rat rasgele dört gruba ayrıldı. Birinci grup kontrol grubu olarak seçildi ve intra peritoneal serum fizyolojik verildi; İkinci gruba intraperitoneal endotoksin; Üçüncü gruba intraperitoneal ornitin ve serum fizyolojik, dördüncü gruba endotoksin ve ornitin verildi. Bulgular: İkinci ve dördüncü grup ratların stria vaskülarislerinde konjeyon, supepitelyal ödem ve modiolusun membranöz labirent bölümünde lökosit ve plazma hücre infiltrasyonu gözlendi. Meatus akustikus internusta yer alan vestibulokoklear ve fasiyal sinirlerde enflamasyon saptanmadı. Gruplar arasında plazma nitrit/nitrat ve MDA seviyelerini karşılaştırdığımızda, birinci ile ikinci, birinci ile üçüncü, birinci ile dördüncü grup arasında anlamlı bir fark bulunamadı. İmmünohistokimyasal olarak hiçbir grupta p53 immünreaktif hücre saptanmadı. Sonuç: L- ornitin rat kokleasındaki septik bulguları düzeltememektedir.

Anahtar Kelimeler: İçkulak immünitesi, labirentit, sepsis, endotoksin, işitme kaybı, endotoksemi.

INTRODUCTION

The prime initiatiator of gram-negative bacterial sepsis is endotoxin, a lipopolysaccharide (LPS) component of the bacterial outer membrane, which induces strong inflammation and cell infiltrations in the cochlea (1, 2). It has been shown that cytokines, released during endotoxemia, can induce nitric oxide synthase (iNOS) in cells which in turn causes production of nitric oxide (NO) in great amounts (3, 4). NO has been demonstrated as an inducer of apoptosis for several years (5,6). During NO mediated apoptosis, as a tumor suppressor gene product, p53 has been shown to accumulate in macrophages (7). Additionally, numerous noxious stimuli such as oncogen activation, hypoxia, hiperoxia, temperature changes, inflamation and DNA damage activate p53 via different mechanism. The activation of p53 due to overproduction of NO during LPS-induced endotoxemia increases cell apoptosis in parenchymall tissues (8). Consequently, their functions may be disconcerted. The aim of therapeutic strategies is to inhibit iNOS or scavenging NO radicals with chemical moieties (1). It was reported that L- ornithine prevented overproduction of NO to inhibit transport of Larginin (9). In order to detect the levels of NO radicals indirectly and oxidative states of organism, the blood levels of malondialdehyde (MDA), nitrate

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and nitrite may be measured. The lower blood levels of MDA, nitrite and nitrate indicate the sufficient antioxidant effect of L-ornithine via decreasing overproduction of NO. In sepsis due to the overproduction of NO from L-arginine, there is an extra demand of this amino acid, which is carried by the same transport system that can be inhibited by Lornithine (1,9). Therefore, we hypothesized that if this transporter was blocked by ornithine, the massive formation of NO could be prevented in sepsis. In the present study, we aimed to investigate the relationship between ornithine treatment, p53 expression and the histopathologic changes in the inner ear of rats in endotoxemia induced by lipopolysaccharide.

MATERIALS AND METHODS

The experiments described in this article were performed in adherence to institutional animal care and study guidelines on the use of experimental animals. This experimental protocol was approved by the ethic committee of our institute. Twentyfour healthy male Wistar rats, weighing 220 – 300 g, were housed at constant temperature with 14/10 h periods of light and dark exposure, respectively.

Experimental protocol:

The rats were randomly divided into 4 groups. The first group, served as control, received intraperitoneal saline (1ml/kg, n=6); the second group, received intraperitoneal endotoxin (Escherchia coli lipopolysacchardie 055: B5 10 mg kg-1 Sigma St Louis, Missouri, n=6); the third group, intraperitoneal saline and L- ornithine (500 mg kg-1, O-2375, Sigma, St Louis, Missouri, n=6), and the fourth group received intraperitoneal L- ornithine (500 mg kg-1) 10 minutes before endotoxin administration (n=6).

Histopathology

Six hours later, rats were anaesthetized with intramuscular ketamine 50 mg kg-1 and xylazine 7 mg kg-1 and animals were sacrificed. The temporal bones were dissected and immersed in the fixative at 4 °C overnight. After decalcification with 10 % ethylenediaminetetraacetic acid, they were processed for paraffin embedding. Sections (4 mm) were cut at 40 mm intervals (midmodiolar hemisection) and stained with hematoxylin and eosin. To assess the severity of inflammation on inner ear cells, the of including number inflammatory cells, polymorphonuclear (PMN) cells, lymphocytes and macrophages were counted (Olympus Bx 50, Japan).

Immunohistochemical staining procedure

The distribution of p53 positive cells were examined by the avidin–biotin-peroxidase

immunohistochemical method. Sections were cut in 5 m thickness and placed onto poly-L-lysine-coated slides. They were dried at room temperature overnight. Following this, sections were deparaffinized in xylene through ethanol to phosphate-buffered saline (PBS; pH 7.2). Endogen peroxidase activity was inhibited using 0.3 % solution of H2O2 in pH 7.5 PBS. Next, the sections were incubated 2 hours with p53 mouse antihuman monoclonal kit (clone DO-7, DAKO, 1:75). Following the first incubation, the sections were incubated for 30 minutes in avidin-biotin-complex peroxidase kit (Labvision Corporation, Fremont, USA). Finally the sections were counter-stained with Mayer's hematoxylen. The number and localization of p53 immunoreactive cells were evaluated in the cochlea.

Detection of plasma nitrate and malondialdehyde levels :

Blood samples were collected from each rat by cardiac puncture. We detected NO via nitrate which presents in the sample. Nitrate was reduced to nitrite by nitrate reductase and the formed nitrite was put to react with sulpanilamide and N-(1-naphtyl)ethylenediamine to give a red-violet diazo dye. The diazo dye was measured on the basis of its absorbance in the visible range at 550 nm (Nitric Oxide Colormetric Assay, Boehringer Manheim, Cat. No. 1756281, Mannheim, Germany).

The levels of serum malondialdehyde (MDA) were measured spectrophotometrically by the method described by Yagi [11]. The final results were expressed as nmol of MDA formed per milliliter of serum. The differences between four groups were evaluated by "t-test" and p< 0.05 was accepted as significant

RESULTS

Histopathology

Congestion and subepitelial edema were observed in stria vascularis, and also infiltration of leukocyte and plasma cell infiltration in perimodiolar region of membraneuos labyrinth of second and fourth groups rats (Figure 1, 2 and 3). Inflammation was not detected in the parts of vestibulocochlear and facial nerves in meatus acousticus internus (Figure 4). The degeneration or lost of hairy cells of cochlea was not observed in any section prepared from group 2 and 4 (Figure 5). Ornithine did not reduce the harmful effect of sepsis on rats from group 4.

Immunohistochemical staining

p53 immunoreactive cells were not detected in any sections of the inner ear.





Figure 1. The normal appearance of cochleary section of control group



Figure 2. Congestion (narrow arrow) and subepithelial severe edema (wide arrowhead) in stria vascularis of group 2 and 4. (H.E. x40)



Figure 3. Periosteal intensive inflammation and neutrophil leukocyt infiltration in cochlea of groups 2 and 4. (H.E. x20)

Plasma nitrate/nitrite and MDA levels

The mean plasma nitrite/nitrate and MDA levels were shown in table 1 (Table 1). When we compared the plasma levels of nitrite/nitrate and MDA among groups, we found statistically significant differences: between groups 1 and 2 (MDA:p<0.001, nitrite: p>0.05, nitrate: p<0.05), groups 1 and 3 (MDA: p<0.05, nitrite: p>0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitrate: p<0.05, nitra

nitrite: p>0.05, nitrate: p<0.05), groups 2 and 3 (MDA: p<0.001, nitrite: p<0.05, nitrate: p<0.001), groups 3 and 4 (MDA: p<0.05, nitrite: p>0.05, nitrate: p>0.05, nitrate: p<0.001). However, no significant difference was found in the comparison of groups 2 and 4 (MDA: p>0.05, nitrite: p>0.05, nitrite: p>0.05).



Figure 4. Normal appearance of meatus acousticuc internus and its containing in gruops 2 and 4 (H.E. x20)



Figure 5. View of normal appearance of tectorial membrane (wide arrow), outer hair cells (narrow arrows) and basilar membrane

Group	MDA (ng/ml)	Nitrite (µmol/ml)	Nitrate (µmol/ml)
Group 1	2.13 ± 0.50	11.36 ± 5.09	4.35 ± 1.68
Group 2	5.01 ± 0.95	14.11 ± 3.64	8.00 ± 1.25
Group 3	1.24 ± 0.22	7.21 ± 4.53	1.45 ± 0.97
Group 4	4.05 ± 1.77	13.76 ± 6.13	7.21 ± 1.78
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Table 1. The mean±standart deviation of serum levels ofnitrite/nitrate and MDA in groups.

DISCUSSION

In the present study, despite inflammation and edema commonly situated in modiolus, spiral ligament and stria vascularis, the degeneration or lost of hairy cell were not observed. The p53 immunoreactive cell was not detected in any section of the inner ear from animals of the second and the fourth group. This condition clearly supported the existence of blood-labyrinth barier in stria vascularis



which protected the organ of corti against inflammation.

The inner ear is enclosed with a bony capsule and lacks lymphatic drainage. The labyrinth is separated from the bloodstream by a barrier that helps to maintain the electrolytic features of the cochlear environment. This blood-labyrinth barrier presents tight junctions in the stria vascularis. It plays an important role in protection of cochlea during meningitis induced by LPS (11-13). Some cytokines, such as TNF-a, interleukin-1 (IL-1), interferon-g (IFN-g), and various colony-stimulating factors are produced when there is a contact between LPS and host defense cells. During LPS induced endotoxemia, excessive NO formation by iNOS increases the expression of inflammatory markers. Massive intraor extracellular NO generation by iNOS has been suggested to cause apoptosis (5). Cytokine induced NO generation causes appereance of typical apoptotic features, DNA fragmentation, nuclear condensation and apoptotic body formation (6). Tumor supressor gene product p53 has been shown to accumulate during NO mediated apoptosis in RAW 264.7 macrophages (5).

On the other hand, Suzuki et al (14) showed that the epithelia of the spiral modiolar vein (SMV) and collecting venule show an increase in expression of intercellular adhesion molecule-1 (ICAM-1) during inflammation. When the inflammatory stimuli causes DNA damage, the expression of p53 increases. There are two major functions of p53, repairing of DNA by stopping cell cycle, and leading cells to apoptosis. In our study, the p53 immunoreactive cells were not detected immunohistochemically. Hence we thought that the severity of cochlear inflammation either was not sufficient or blood–labyrinth barier protected the inner ear from endotoxin to induce DNA damage in the inner ear cells of rats.

Because NO has been detected in the circulation and cerebrospinal fluid of septic patients, it has become the target of therapeutic strategies aimed at inhibiting iNOS or scavenging NO radicals with chemical moieties (1). It was reported that Lornithine prevented overproduction of NO by inhibiting transport of L-arginin (9). Although Lornithine is assumed to be a scavenging of NO, we could not observe this effect of L-ornithine biochemically and histopathologically. Furthermore, plasma nitrite/nitrate levels were reduced by the treatment of L-ornithine but this reduction was not significant statistically (p>0.05). This condition indicated either that the L-ornithine failed to prevent L-arginine transport to the cell or intracellular Larginine storage is well preserved and thus not affected by the inhibition of L-ornithine in the endotoxemia.

CONCLUSION

Although inflammation is observed in different degrees especially in the stria vascularis and perimodiolar region, it does not spread to inner ear through the vestibulocochlear and facial nerve. The severity of cochlear inflammation was not sufficient to induce DNA damage in the inner ear cell. Additionally, it seems unlikely that L-ornithine at this dose can improve the manifestations of inflammation of inner ear during this type of endotoxemia.

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